



Mission pour les initiatives
transverses et interdisciplinaires
(MITI)



Journée thématique

Fonctionnalisation de sonde et analyses de données

Toulouse, 29 novembre 2023

Interaction and regulation in redox partner proteins: electrochemical-STM and AFM-force spectroscopy

Marina I. Giannotti

Nanoprobes & Nanoswitches

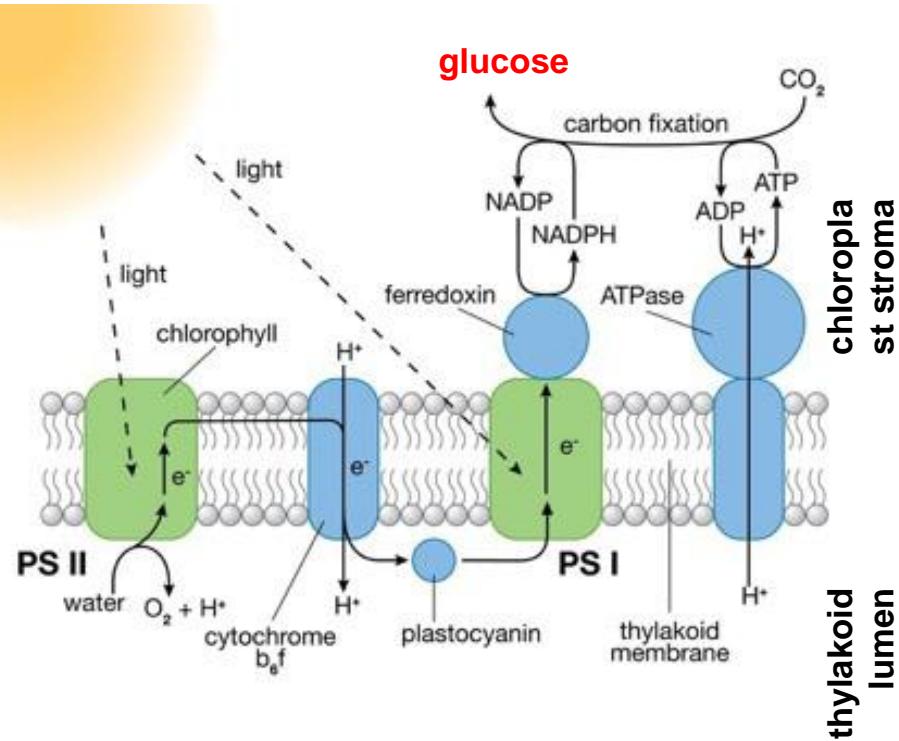
Institute for Bioengineering of Catalonia **IBEC**

Pau Gorostiza



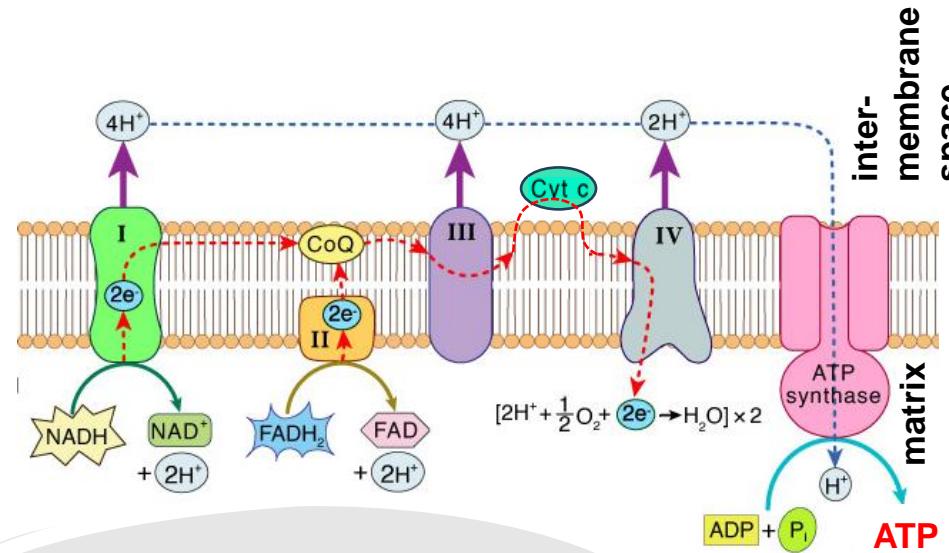
CHARGE TRANSPORT IN PHOTOSYNTHETIC AND RESPIRATORY COMPLEXES protein-mediated electron transport (ET)

PLANT PHOTOSYNTHETIC CHAIN



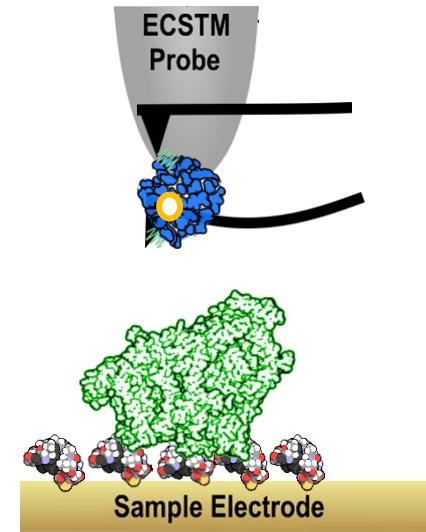
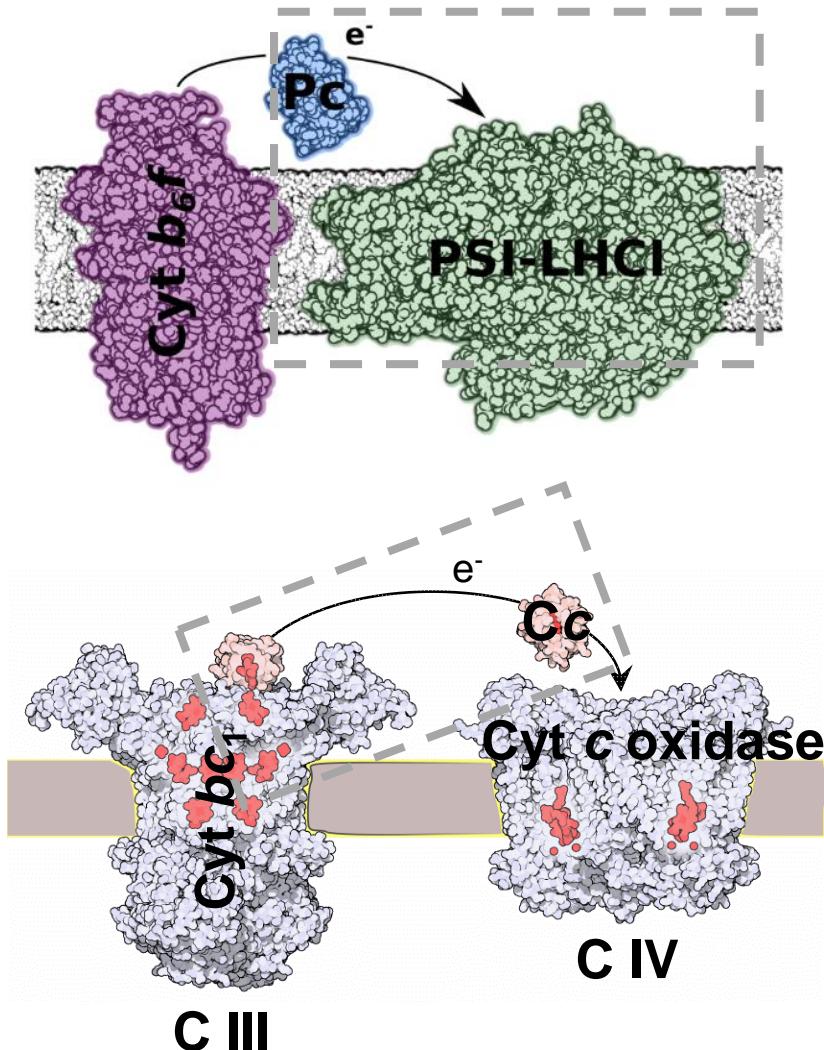
sustained and efficient electron flow

MITOCHONDRIAL RESPIRATORY CHAIN

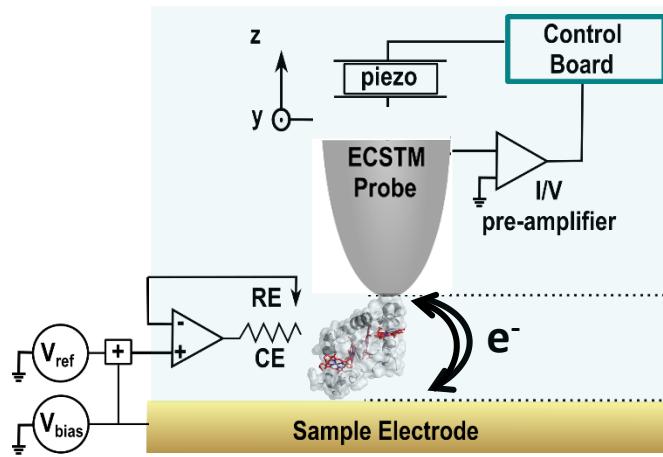


reorganization energy ↓
turn-over ↑

inter-biomolecular interactions at the nanoscale



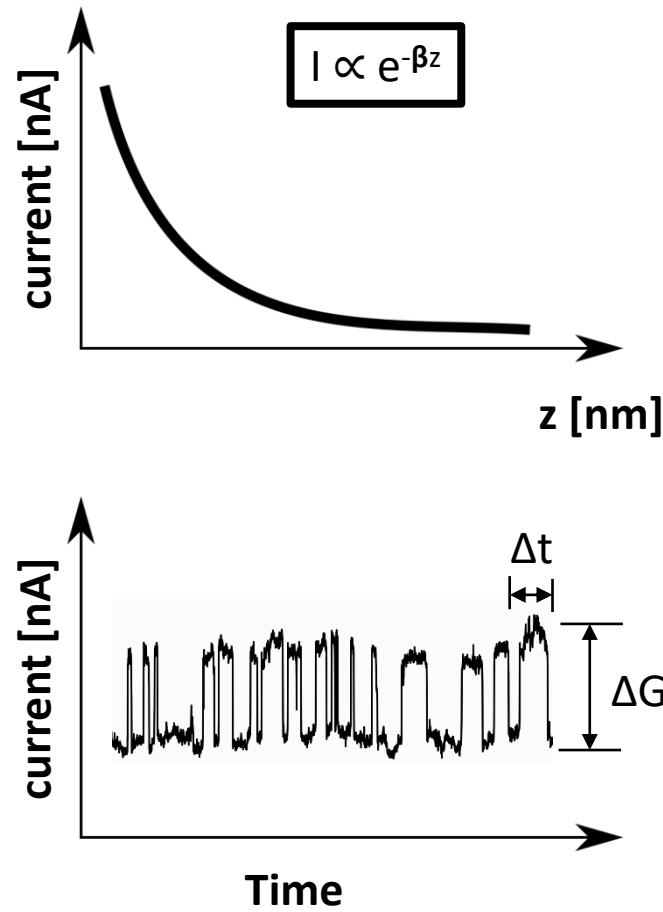
Electrochemical STM



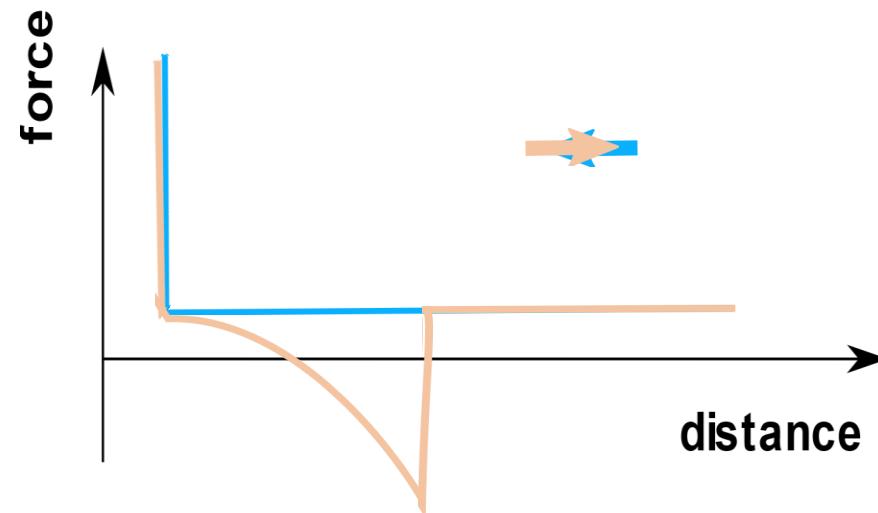
$$U_{\text{bias}} = U_{\text{probe}} - U_{\text{sample}}$$

$$U_{\text{bias}} < 0 \quad I > 0$$

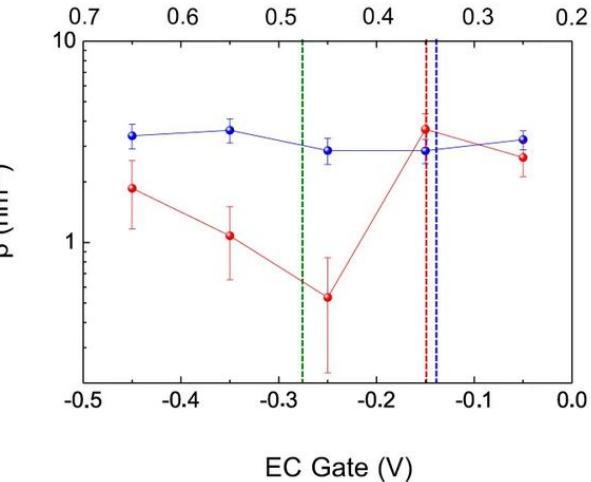
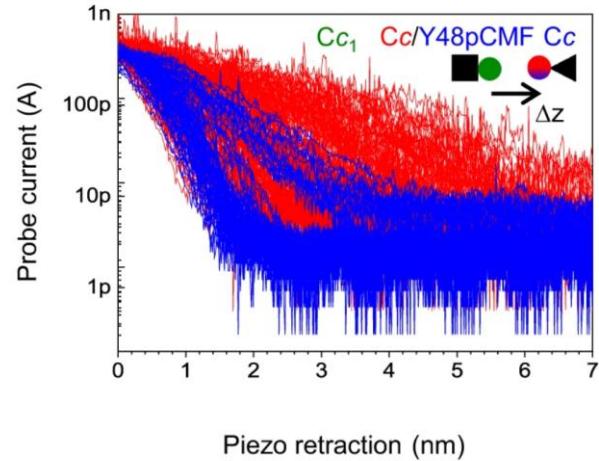
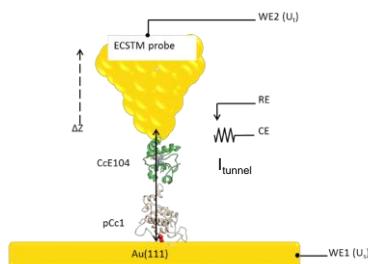
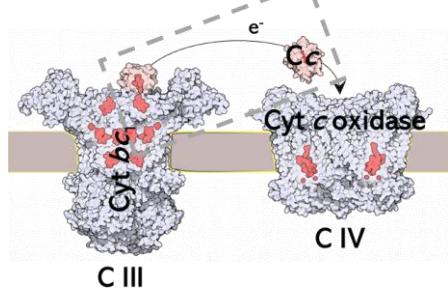
$$U_{\text{bias}} > 0 \quad I < 0$$



AFM force spectroscopy

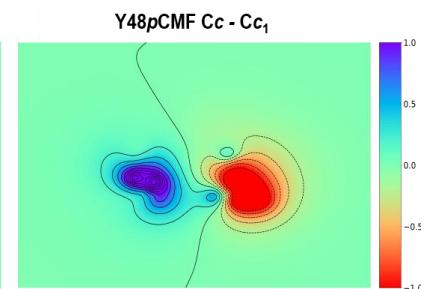
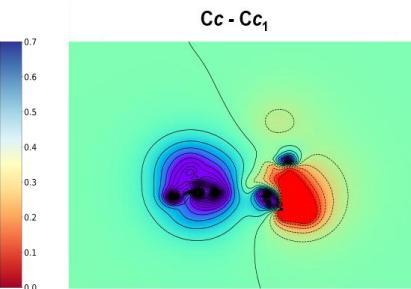
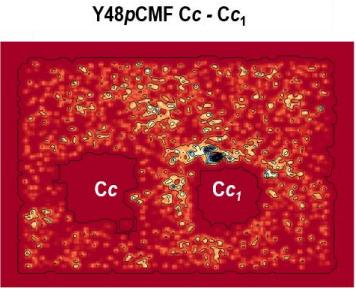
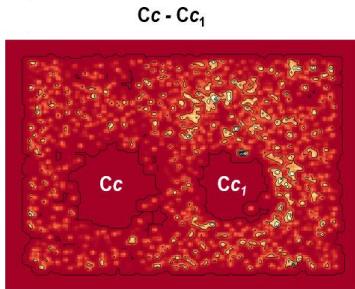


Cytochrome c (Cc)– cytochrome c_1 (Cc_1)

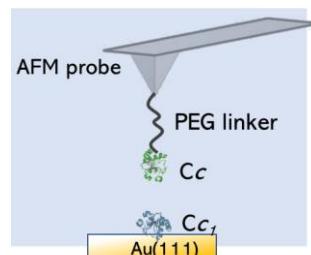
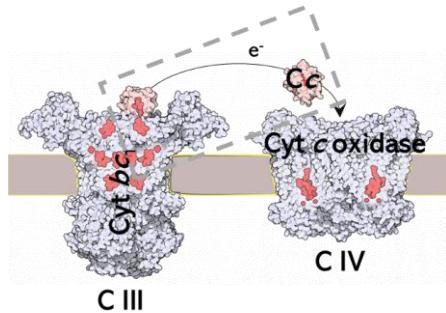


Lagunas et al. Nat. Commun. 2018

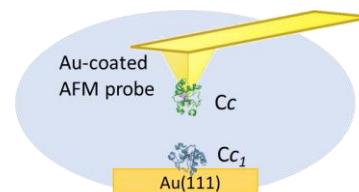
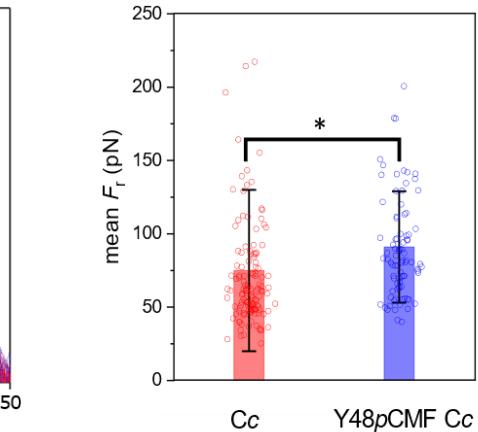
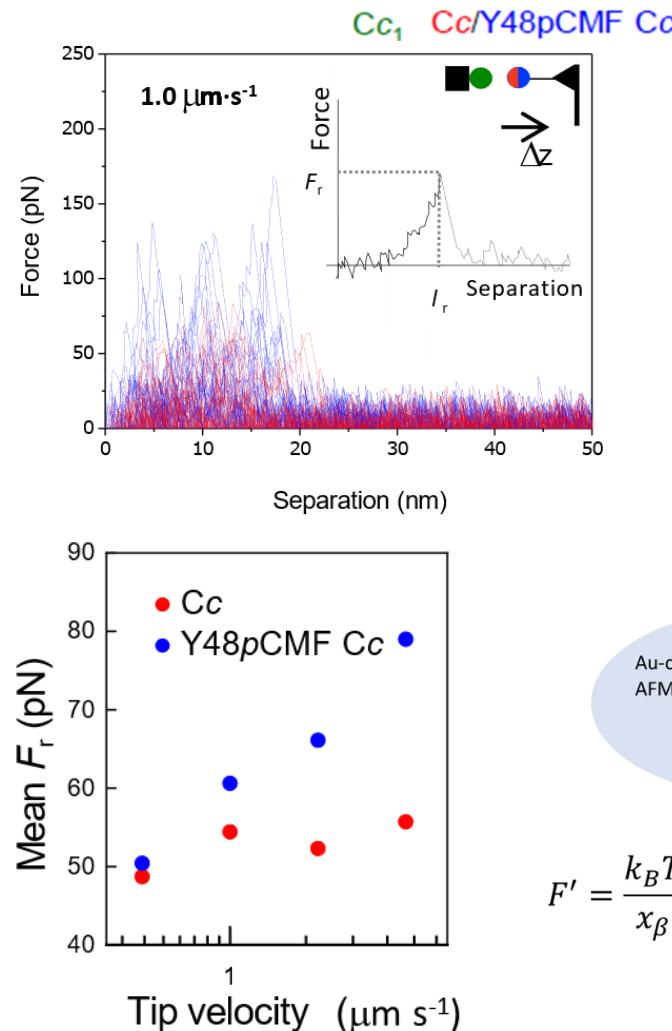
Gomila et al. Nat. Commun. 2022



Cytochrome c (Cc)– cytochrome c_1 (Cc_1)

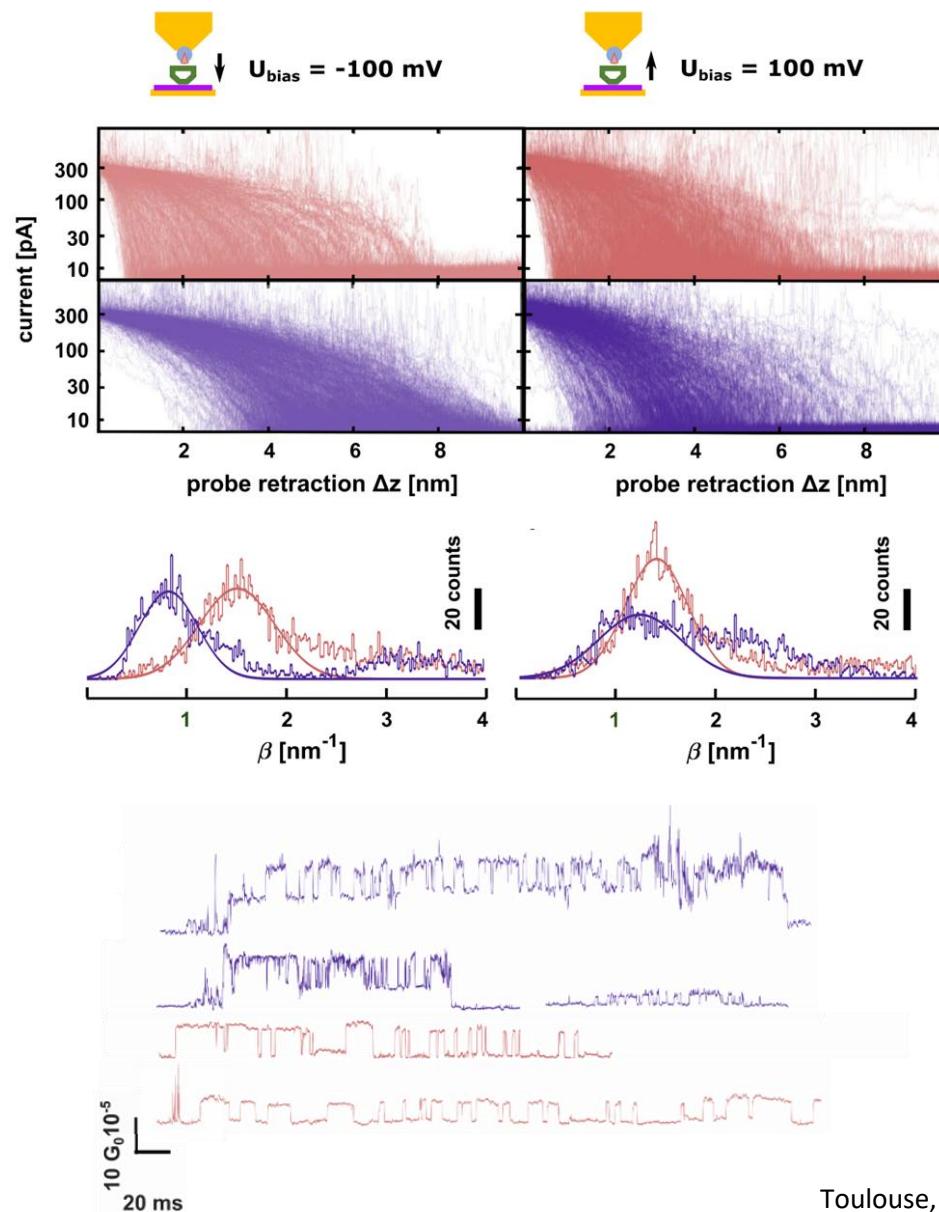
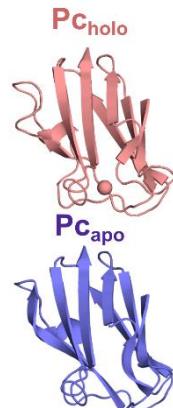
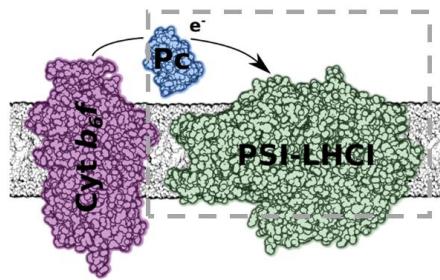


Gomila et al. Nat. Commun. 2022



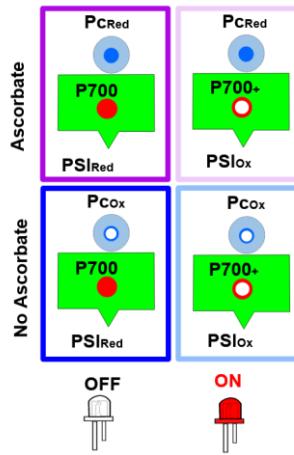
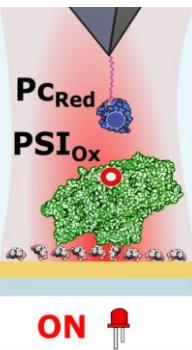
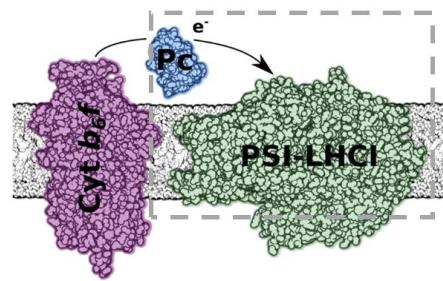
$$F' = \frac{k_B T}{x_\beta} \cdot \left[(\ln r) + \ln \frac{x_\beta}{k_{off} \cdot k_B T} \right]$$

PSI – Plastocyanin (Pc)

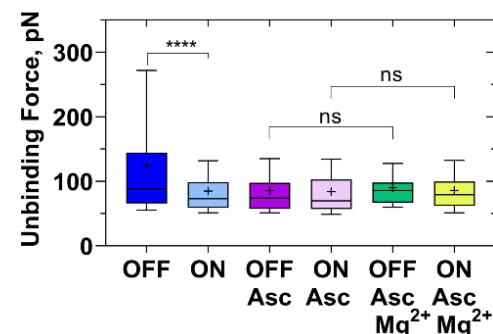
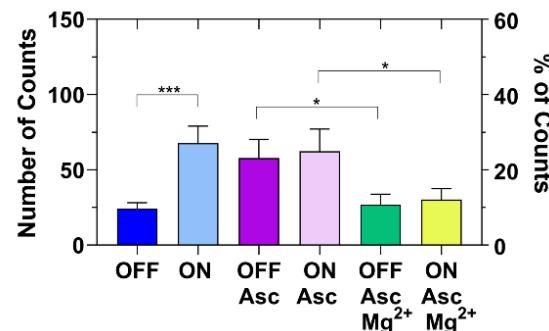
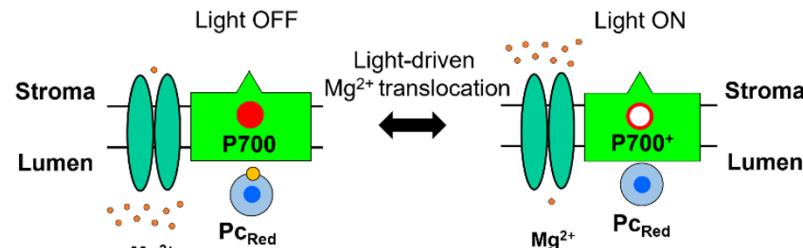
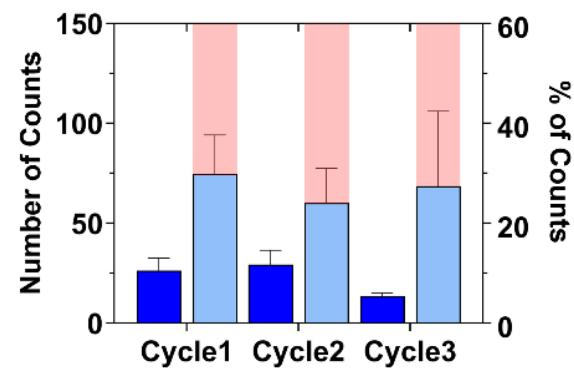
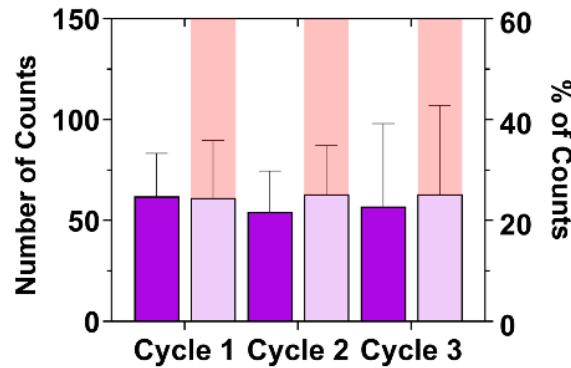


López-Ortíz et al. ACS Nano 2023

PSI – Plastocyanin (Pc)



Zamora et al. ACS Nano 2022



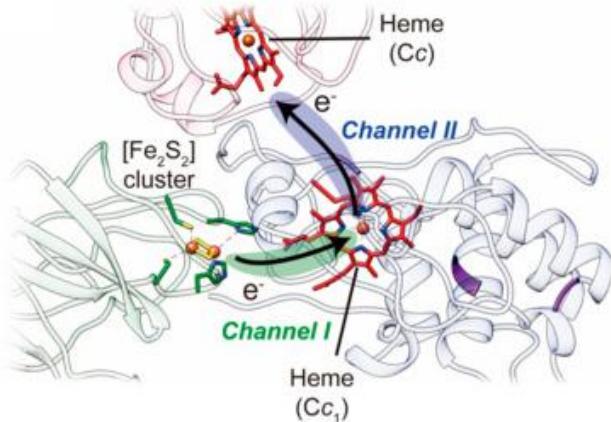
Main observations

- ✓ long-range ET in PSI-Pc and Cc-Cc₁ in aqueous solution, ion depletion conduit (ECSTM on oriented protein cognates)
 - ✓ charge exchange distance and conductance is modulated by the cooper ion in Pc (PSI-Pc)
 - ✓ Phosphorylation impairs ET (Cc-Cc₁)

binding as a regulatory mechanism for ET rates
rather than a necessary condition for ET

- ✓ specific but **weak binding**, necessary for high efficiency and turn-over rate (AFM-FS on oriented protein cognates)
 - ✓ regulated by redox state, Mg²⁺ ions in solution (PSI-Pc),
 - ✓ regulated by chemical modifications (PTM) like phosphorylation (Cc-Cc₁)

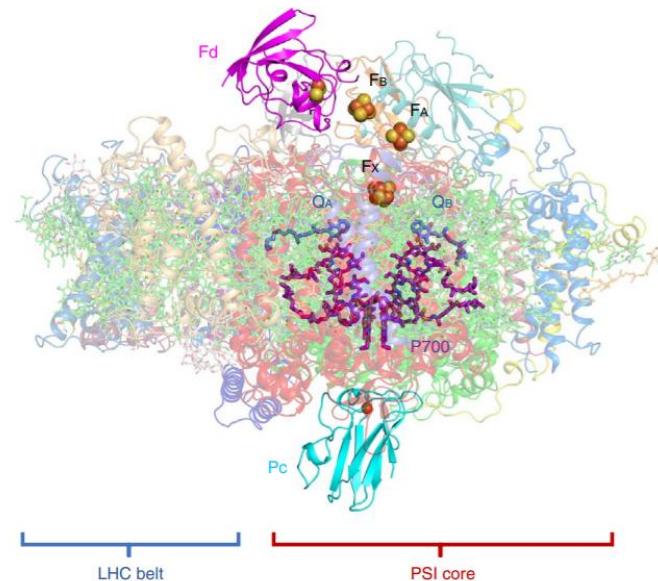
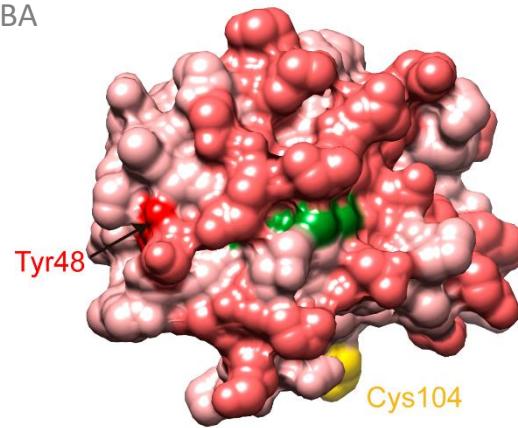
Protein immobilized with right orientation



Pérez-Mejías et al. BBA
Bioenergetics 2020

Cc-SH
E104C mutant

Cytochrome c



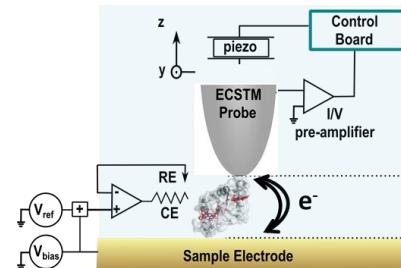
Casy et al. Nature Plants 2020

Pc-SH
mutant with three extra
residues at the C-terminal:
Thr-Cys-Gly

Plastocyanin

Electrochemical STM

Au ECSTM Probes. Functionalization with Pc_{apo} /holo



Mechanically sharpen
the Au wire

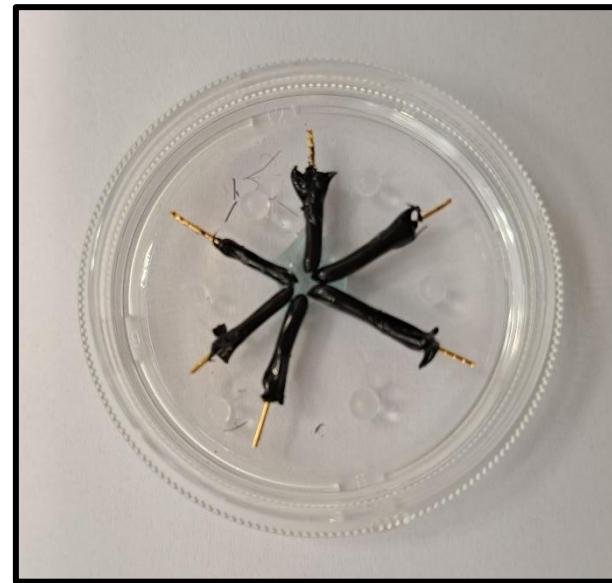
Coat with Apiezon
wax

Au probes placed
in a PDMS-coated
Petri dish

1 hour incubation at
4°C of purified Pc_{apo}

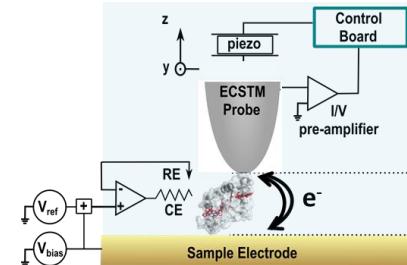


~120°C



Electrochemical STM

Au ECSTM Probes. Functionalization with Pc_{apo} /holo



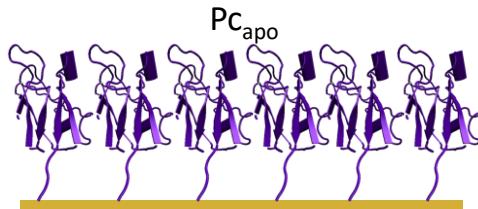
Rinse with MES the unbound Pc_{apo}

1 hour incubation at 4°C of CuSO_4 – MES solution

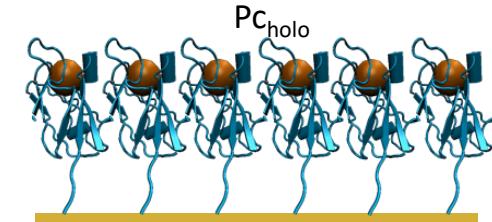
Rinse with MES the unbound Cu

Pc_{holo} is ready for experiments

Pc_{apo} is ready for experiments



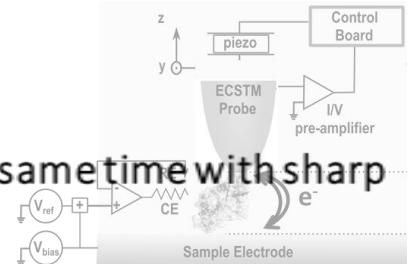
Au wire



Au wire

Au ECSTM PROBES FUNCTIONALIZATION:

1. Mechanically sharpen the gold wire: cut the Au wire and pull and cut at the same time with sharp hand-held steel wire cutter to obtain the tip
2. Coat twice the Au wire with Apiezon wax heated at ~120°C with a home-built heat-coating applicator
3. Place 6 probes looking at each other in a polydimethylsiloxane (PDMS)-coated Petri dish
4. Place 100 µL drop of purified protein (Pc_{apo}) solution soaking all probe tips and incubate for 1 hour at 4°C
5. Put drops of water around the probes to maintain humidity
6. Extract the protein (Pc_{apo}) drop and rinse with protein buffer to eliminate the unbound protein
7. For Pc_{holo} incubate 1 hour more at 4°C with a solution of MES 5 mM pH = 5.5 with 100 mM of $CuSO_4$ and rinse again with MES after to eliminate the unbound Cu^{2+}



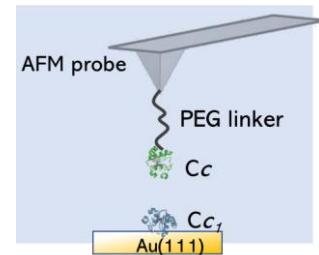
Lagunas et al. Nat. Commun. 2018

Gomila et al. Nat. Commun. 2022

López-Ortíz et al. ACS Nano 2023

AFM force spectroscopy

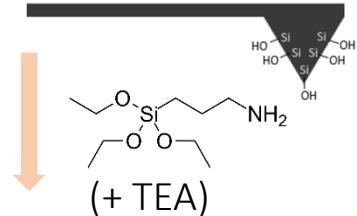
Si/Si₃N₄ AFM probes. Functionalization with PEG-protein



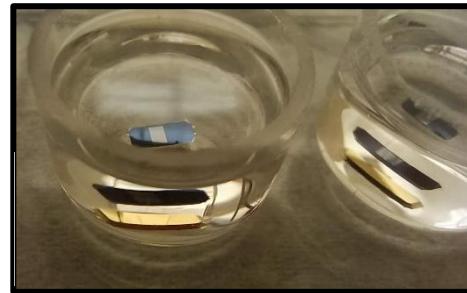
Surface activation
O₂/UV



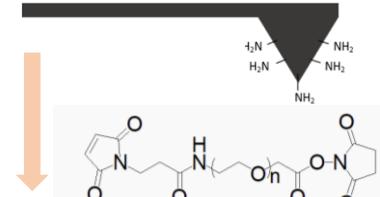
APTES
functionalization
(vapor phase) ON



Rinse chloroform
and ethanol, dry
N₂ (can be stored)

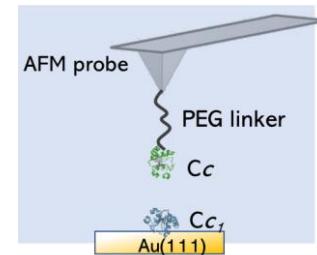


React with
heterobifunctional PEG
(+ TEA) /chloroform 1,5 h

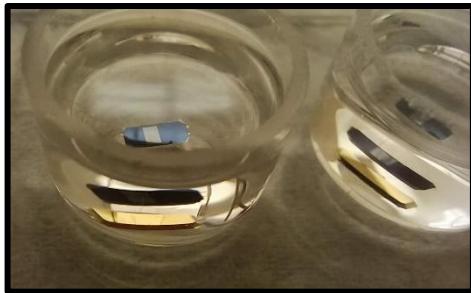


AFM force spectroscopy

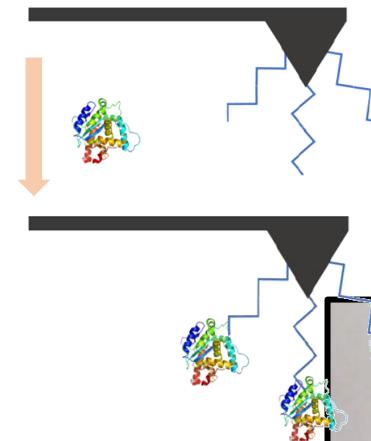
Si/Si₃N₄ AFM probes. Functionalization with PEG-protein



Rinse chloroform
and ethanol and
MilliQ Water, dry N₂



1 hour incubation at
4°C of purified Pc_{apo}



Rinse with buffer

1 hour incubation at
4°C of CuSO₄ – MES
solution

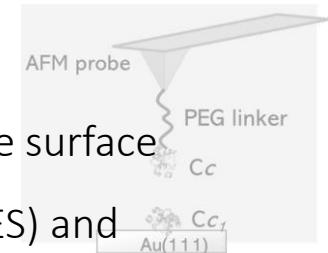
Rinse with MES the
unbound Cu

Pc_{holo} is ready for
experiments



Si/Si₂N₃ AFM PROBES FUNCTIONALIZATION:

1. Treat 15 min with a UV/Ozone ProCleaner (BioForce Nanosciences) to activate surface
2. Amino-functionalize in an atmosphere of 3-aminopropyltriethoxysilane (APTES) and triethylamine (TEA). Inside the desiccator previously flooded with Ar or N₂. Two containers with 45 µL APTES and 15 µL TEA, separately, close to the probes (on clean inert surface), vacuum on for 5 min. After 1.5 h, remove APTES and TEA containers, and vacuum is restored. Overnight, RT
3. Rinse chloroform (2x) and ethanol (2x) and dry under gentle N₂ flow. When not used immediately, they can be stored in a desiccator under Ar
4. Place in glass wells with a 0.5 mL solution 1 mg/mL heterobifunctional PEG (Mal-PEG27-NHS) and 5 µL of TEA in chloroform, 1.5 h, RT. Rinse with chloroform, ethanol and MilliQ water
5. Place 3 probes looking at each in a PDMS-coated Petri dish. Place 25 µL drop of purified protein solution (1 mg/mL) over all probe tips and incubate for 1 hour at 4°C
6. Rinse with protein buffer to eliminate the unbound protein



Acknowledgements

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